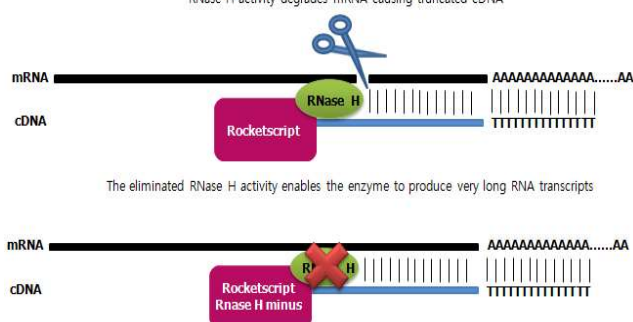


[Cat. No.] **K-2249**

### Introduction

AccuPower® RocketScript™ RT Master Mix, RNase H Minus uses RocketScript™ Reverse Transcriptase (RTase), RNase H Minus independently developed by BIONEER. RocketScript™ RTase, RNase H Minus is a recombinant *M-MLV* RTase with enhanced thermostability up to 70°C, as well as eliminated RNase H activity. It is highly efficient in producing full-length cDNA from long RNA transcripts up to 12.5 kb. In addition, it can synthesize cDNA even with small amounts of 1 pg of total RNA. This product is a ready-to-use mixture for cDNA synthesis including RocketScript™ RTase, RNase H Minus, reaction buffer, DTT, dNTPs, and RNase inhibitor. It simplifies preparation of reverse transcription reaction mixture by adding template RNA, primers, and nuclease-free water without any extra process.

RNase H activity degrades mRNA causing truncated cDNA



The eliminated RNase H activity enables the enzyme to produce very long RNA transcripts

**Figure 1. Elimination of RNase H activity.** Eliminated RNase H activity from RTase complex enables the RTase to produce longer RNA transcripts, up to 12.5 kb, compared with original RocketScript™ RTase itself.

### Applications

- First-strand synthesis of cDNA from RNA molecules (RT)
- Random priming reaction
- cDNA library construction, Probe labeling
- mRNA 5'-end mapping by primer extension analysis
- PCR, Real-time PCR

### Features & Benefits

- Rapid synthesis (9 kb synthesis possible with 10 minutes RT reaction): Time-efficient and economical cDNA synthesis even with secondary structure RNA can be possible by using RocketScript™ RTase due to excellent thermal stability.
- No RNase H activity: Useful for cDNA synthesis up to 12.5 kb from long target RNA using enzymes developed by proprietary genetic engineering technology.
- Sensitivity: Efficient cDNA synthesis even with small amounts of 1 pg of human total RNA.

### Components

Components	Amount
2X Master Mix	1 ml
Oligo dT <sub>20</sub> (100 pmol/μl)	100 μl
Oligo dN <sub>6</sub> (100 pmol/μl)	100 μl

### Composition

2X Master Mix	Concentration
RocketScript™ Reverse Transcriptase, RNase H Minus	200 U
5X Reaction buffer	1X
DTT	0.25 mM
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 μM
RNase inhibitors	1 U

### Specifications

RocketScript™ Reverse Transcriptase, RNase H Minus	
DNase activity	No
RNase activity	No
RNase H activity	No
Fragment size	Up to 12.5 kb

### Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

### Online Resources



Korean



English

Visit our **product page** for additional information and protocols

### Ordering Information

Description	Cat. No.
1 ml of 2X Master Mix solution	1 ml x 1 ea K-2249

\* **Note:** For Master Mix products, primers (Oligo dT<sub>20</sub> and Oligo dN<sub>6</sub>) are provided in a separate tube.





### Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

### Explanation of Symbols

Batch Code	Biological Risks	Catalog Number	Caution
Consult Instructions For Use	Contains Sufficient for <n> tests	Do not Re-use	Manufacturer
Research Use Only	Temperature Limitation	Use-by Date	

**Experimental Procedures**

Steps		Procedure Details																																				
1	 <b>Thaw reagents</b>	<p>1. Thaw <i>AccuPower® RocketScript™</i> RT Master Mix, RNase H Minus on ice and mix thoroughly before use. Then, briefly spin down components.</p> <p>2. Dispense appropriate volumes of <i>AccuPower® RocketScript™</i> RT Master Mix, RNase H Minus into PCR tubes (not provided). Use 10 µl and 25 µl of 2X Master Mix for 20 µl reaction and 50 µl reaction, respectively.</p>																																				
2	 <b>Preparation of reaction mixture</b>	<p>3. Add template RNA and primers into PCR tubes containing <i>AccuPower® RocketScript™</i> RT Master Mix, RNase H Minus, and then fill up nuclease-free water to make a total volume of 20 µl or 50 µl.</p> <ul style="list-style-type: none"> <li>Amount of template RNA and primers</li> </ul> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th colspan="2">Components</th> <th>20 µl reaction</th> <th>50 µl reaction</th> </tr> </thead> <tbody> <tr> <td rowspan="2" style="text-align: center;">Template RNA</td> <td style="text-align: center;">Total RNA</td> <td style="text-align: center;">0.01-5 µg</td> <td style="text-align: center;">0.01-5 µg</td> </tr> <tr> <td style="text-align: center;">Poly(A) RNA</td> <td style="text-align: center;">0.01-5 µg</td> <td style="text-align: center;">0.01-5 µg</td> </tr> <tr> <td rowspan="3" style="text-align: center;">Primers</td> <td style="text-align: center;">Oligo dT</td> <td style="text-align: center;">100 pmol</td> <td style="text-align: center;">250 pmol</td> </tr> <tr> <td style="text-align: center;">Random primer</td> <td style="text-align: center;">100 pmol</td> <td style="text-align: center;">250 pmol</td> </tr> <tr> <td style="text-align: center;">Gene specific primer</td> <td style="text-align: center;">10-50 pmol</td> <td style="text-align: center;">10-50 pmol</td> </tr> </tbody> </table> <p>4. Mix the reaction mixture by pipetting or vortexing, and briefly spin down.</p>	Components		20 µl reaction	50 µl reaction	Template RNA	Total RNA	0.01-5 µg	0.01-5 µg	Poly(A) RNA	0.01-5 µg	0.01-5 µg	Primers	Oligo dT	100 pmol	250 pmol	Random primer	100 pmol	250 pmol	Gene specific primer	10-50 pmol	10-50 pmol															
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3	 <b>cDNA synthesis</b>	<p>5. Perform the reaction under the following conditions.</p> <p>5-1. Example 1</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th rowspan="2">Step</th> <th colspan="3">Temperature</th> <th rowspan="2">Gene specific</th> <th rowspan="2">Time</th> </tr> <tr> <th>dN<sub>6</sub></th> <th>dN<sub>12</sub></th> <th>dT<sub>20</sub></th> </tr> </thead> <tbody> <tr> <td>Primer annealing</td> <td style="text-align: center;">15°C</td> <td style="text-align: center;">30°C</td> <td style="text-align: center;">37°C</td> <td style="text-align: center;">T<sub>m</sub> of primers</td> <td style="text-align: center;">10 min</td> </tr> <tr> <td>cDNA synthesis</td> <td></td> <td></td> <td style="text-align: center;">50°C</td> <td></td> <td style="text-align: center;">30 min</td> </tr> <tr> <td>Heat inactivation</td> <td></td> <td></td> <td style="text-align: center;">95°C</td> <td></td> <td style="text-align: center;">5 min</td> </tr> </tbody> </table> <p>5-2. Example 2</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> </tr> </thead> <tbody> <tr> <td>cDNA synthesis</td> <td style="text-align: center;">42-70°C</td> <td style="text-align: center;">1 hr</td> </tr> <tr> <td>Heat inactivation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> </tr> </tbody> </table> <p>* <b>Note:</b> For difficult or high GC-content templates, perform cDNA synthesis at 55°C.</p> <p>6. After the reaction, maintain the reaction mixture at 4°C. The samples can be stored at -20°C until use.</p>	Step	Temperature			Gene specific	Time	dN <sub>6</sub>	dN <sub>12</sub>	dT <sub>20</sub>	Primer annealing	15°C	30°C	37°C	T <sub>m</sub> of primers	10 min	cDNA synthesis			50°C		30 min	Heat inactivation			95°C		5 min	Step	Temperature	Time	cDNA synthesis	42-70°C	1 hr	Heat inactivation	95°C	5 min
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	 <b>Option</b>	<ul style="list-style-type: none"> <li>If PCR is needed, transfer 2-5 µl of reaction mixture (synthesized cDNA) into <i>AccuPower®</i> PCR PreMix tubes (Cat. No. K-2012, not provided), and perform the reaction under the following conditions.</li> </ul> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">20 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">45-65°C</td> <td style="text-align: center;">20 sec</td> <td style="text-align: center;">25-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">0.5-1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">3-5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* <b>Note:</b> For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	20 sec		Annealing	45-65°C	20 sec	25-35 cycles	Extension	72°C	0.5-1 min/kb		Final extension	72°C	3-5 min	1 cycle												
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