

[Cat. No.] Please refer to the Ordering Information

Introduction

AccuPower® RT PreMix contains vacuum-dried components essential for cDNA synthesis including M-MLV Reverse Transcriptase, reaction buffer, and RNase inhibitor. It simplifies preparation of reverse transcription reaction mixture by adding template RNA and primers without any extra process. Furthermore, by using the RNase H⁺ of M-MLV Reverse Transcriptase, template RNA is removed after the cDNA synthesis to minimize the effect of residual template RNA in PCR.

Applications

- First-strand synthesis of cDNA from RNA molecules (RT)
- Random priming reaction
- cDNA library construction
- Probe labeling
- mRNA 5'-end mapping by primer extension analysis
- Real-time PCR

Features & Benefits

- User-friendly: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform cDNA synthesis by adding template RNA and primers.
- Stability: Included stabilizer and RNase inhibitor provides high resistance to degradation.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration		
M-MLV Reverse Transcriptase	200 U		
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 μM		
5X Reaction buffer	1X		
DTT	0.25 mM		
Stabilizer	1X		

Specifications

M-MLV Reverse Transcriptase				
DNase activity	No			
RNase activity	No			
Fragment size	Up to 9 kb			

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label

Online Resources





Visit our product page for additional information and protocols

Ordering Information

Desci	Cat. No.		
	96 tubes	20 µl/rxn	K-2041
0.2 ml thin-wall 8-tube strips		50 μl/rxn (with dye)	K-2043
with attached cap	480 tubes	20 µl/rxn	K-2041-B
		50 μl/rxn (with dye)	K-2043-B
0.5 ml thin-wall tubes	100 tubes	20 μl/rxn (with dye)	K-2040
with attached cap		50 μl/rxn (with dye)	K-2042
	flat plate	10 µl/rxn	K-2261-1
		20 µl/rxn	K-2261-4
thin wall 06 wall	full-skirted	10 µl/rxn	K-2261-2
thin-wall 96-well	plate	20 µl/rxn	K-2261-5
	semi-skirted plate	10 µl/rxn	K-2261-3
		20 µl/rxn	K-2261-6
	full-skirted plate	5 µl/rxn	K-2082-1
thin-wall 384-well		10 µl/rxn	K-2082-2
		20 µl/rxn	K-2082-3

^{*} Note: The RT series does not contain tracking dyes for electrophoresis, with the exception of some products.

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols





Biological Risks

















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Experimental Procedures

	Steps	Procedure	ocedure Details			
		Mix template RNA and primers in a sterile tube (not provided) indicated as below.				
			Amount of template RNA and primers			
		Com	nponents	20 μl reaction	50 μl reaction	
		Template RNA	Total RNA	0.5-1 μg	1-2 µg	
1			Poly(A) RNA	0.05-0.1 μg	0.1-0.2 μg	
			Oligo dT	100 pmol	200 pmol	
		Primers	Random primer	100 pmol	200 pmol	
	Primer annealing		Gene specific primer	10-30 pmol	20-50 pmol	
		2. Incubate the mixture at 70°C for 5 min and place it on ice directly.				
		 3. Transfer the incubated primer annealing mixture to <i>AccuPower</i>[®] RT PreMix tube, and then fill up the nuclease-free water to make a total volume of 20 μl or 50 μl. 4. Dissolve the vacuum-dried pellet by pipetting or vortexing, and briefly spin down. 				
	ONEER	5. Perform the read	5. Perform the reaction under the following conditions.			
2	cDNA synthesis	Step	Temperature		Time	
		cDNA synthesis	42°0		60 min	
		Heat inactivation	95°C		5 min	
		6. After the reaction, maintain the reaction mixture at 4°C. The samples can be stored at -20°C until use.				
	Option	 If PCR is needed, transfer 2-5 μl of reaction mixture (synthesized cDNA) into AccuPower[®] PCR PreMix tubes (K-2012, not provided), and perform the reaction under the following conditions. 				
		Step	Temperature	Time	Cycles	
		Pre-denaturation	95°C	5 min	1 cycle	
		Denaturation	95°C	20 sec		
		Annealing	45-65°C	20 sec	25-35 cycles	
		Extension	72°C	0.5-1 min/kb		
		Final extension	72°C	3-5 min	1 cycle	
		* Note: For maximum each new template [n yield and specificity, tempera DNA or primers.	tures and cycling times	should be optimized foi	