

[Cat. No.] Please refer to the **Ordering Information**

Introduction

AccuPower® PyroHotStart Taq PCR PreMix is based on a concept chemical interaction between pyrophosphate (PPi) and pyrophosphatase (PPase). DNA polymerase is required Mg²⁺ to activate, but included PPi binds with high affinities to Mg²⁺ resulting in inhibition of polymerase activity. When the temperature rises during denaturation step, Mg-PPi complex is decomposed into 2Pi and Mg²⁺ by PPase. Then, activated DNA polymerase proceed reactions. This prevents the formation of mis-primed products and primer-dimers during the reaction set up process resulting in improved PCR specificity. This product contains vacuum-dried components including *PyroHotStart Taq* DNA Polymerase, dNTPs, reaction buffer, stabilizer, and tracking dye. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, samples can be applied directly on agarose gel for analysis.

Applications

- High specificity PCR
- High sensitivity PCR
- gDNA template PCR
- Low-copy target PCR
- Multiple primer pairs PCR
- cDNA template PCR
- TA cloning

Features & Benefits

- **Specificity:** Minimized non-specific amplification and maximized PCR efficiency by using BIONEER's differentiated PyroHotStart technology.
- **Sensitivity:** Excellent sensitivity and amplification efficiency even with small amounts of DNA.
- **Stability:** Included stabilizer enables delivery at room temperature and provides increased stability compared to solution-type products.
- **User-friendly:** Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- **Reproducibility:** Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
<i>PyroHotStart Taq</i> DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction buffer with 1.5 mM MgCl ₂	1X
Stabilizer and tracking dye	O

Specifications

<i>PyroHotStart Taq</i> DNA Polymerase	
5' to 3' exonuclease activity	Yes
3' to 5' exonuclease activity	No
3'-A overhang	Yes
Fragment size	Up to 5 kb (human)

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

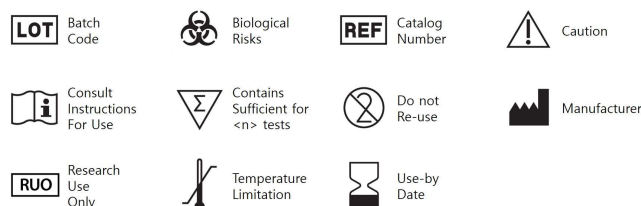
Ordering Information

Description		Cat. No.	
0.2 ml thin-wall 8-tube strips with attached cap	96 tubes	20 µl/rxn	K-2611
	480 tubes	50 µl/rxn	K-2613
		20 µl/rxn	K-2612
		50 µl/rxn	K-2614



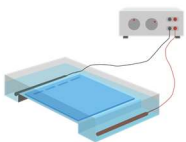

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Experimental Procedures

Steps		Procedure Details																										
1	 Preparation of reaction mixture	<p>1. Add template DNA, primers, and nuclease-free water into <i>AccuPower® PyroHotStart Taq PCR PreMix</i> tubes to make a total volume of 20 µl or 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Components</th> <th style="text-align: center;">20 µl reaction</th> <th colspan="2" style="text-align: center;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA (1-500 ng)</td> <td style="text-align: center;">Variable (1-10 µl)</td> <td colspan="2" style="text-align: center;">Variable (1-25 µl)</td> </tr> <tr> <td>Forward primer (10 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td colspan="2" style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Reverse primer (10 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td colspan="2" style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td style="text-align: center;">Variable</td> <td colspan="2" style="text-align: center;">Variable</td> </tr> <tr> <td>Total volume</td> <td style="text-align: center;">20 µl</td> <td colspan="2" style="text-align: center;">50 µl</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried blue pellet by vortexing or pipetting, and briefly spin down.</p>			Components	20 µl reaction	50 µl reaction		Template DNA (1-500 ng)	Variable (1-10 µl)	Variable (1-25 µl)		Forward primer (10 pmol/µl)	0.5-2 µl	1-5 µl		Reverse primer (10 pmol/µl)	0.5-2 µl	1-5 µl		Nuclease-free water	Variable	Variable		Total volume	20 µl	50 µl	
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2	 Incubate reactions in a thermal cycler	<p>3. Perform the reaction under the following conditions.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">15-30 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">45-65°C</td> <td style="text-align: center;">15-30 sec</td> <td style="text-align: center;">25-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">3-5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table>			Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	15-30 sec		Annealing	45-65°C	15-30 sec	25-35 cycles	Extension	72°C	1 min/kb		Final extension	72°C	3-5 min	1 cycle
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3	 Analyze with gel electrophoresis	<p>4. After the reaction, maintain the reaction mixture at 4-8°C. The samples can be stored at -20°C until use.</p> <p>5. Load 5 µl of samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																										
	 Option	<ul style="list-style-type: none"> If primer's T_m value is more than 65°C or PCR product size is more than 5 kb, follow the conditions as below. <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">30 sec</td> <td></td> </tr> <tr> <td>Annealing/Extension</td> <td style="text-align: center;">68°C</td> <td style="text-align: center;">1 min/kb</td> <td style="text-align: center;">30-35 cycles</td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">3-5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table>			Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	30 sec		Annealing/Extension	68°C	1 min/kb	30-35 cycles	Final extension	72°C	3-5 min	1 cycle				
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