

[Cat. No.] **K-2012-1, K-2016-1**

Introduction

AccuPower® PCR PreMix (with UDG) helps to minimize carryover contamination, which may cause severe problems in clinical diagnosis by using uracil DNA glycosylase (UDG). UDG catalyzes the hydrolysis of N-glycosylic bond between the uracil and sugar. In the following heating at 95°C, contaminants (uracil-containing DNA) are degraded and consequently not amplified. UDG efficiently remove uracil from single-stranded or double-stranded DNA, but from oligomers (6 or fewer). It is not active for targeting RNA or uracil-free DNA. This product contains vacuum-dried components including UDG, *Top* DNA Polymerase, dNTPs with dUTP, reaction buffer, stabilizer, and tracking dye. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, samples can be applied directly on agarose gel for analysis.

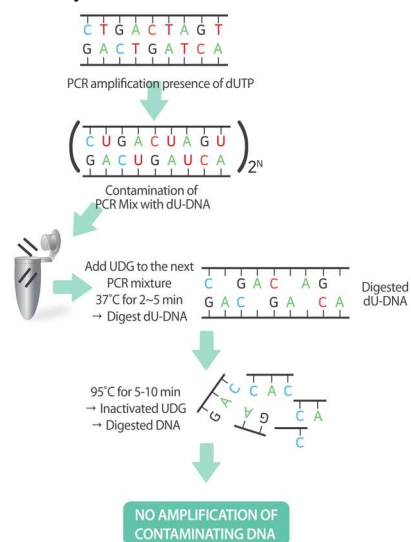


Figure 1. Principle of eliminating contaminants using UDG.

Applications

- Conventional PCR
- Primer extension
- TA cloning
- Gene sequencing
- Molecular diagnosis

Features & Benefits

- Carryover contamination prevention: Minimized false positives caused by a carryover contamination through application of uracil DNA glycosylase system.
- User-friendly: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- Stability: Included stabilizer enables to maintain enzymatic activities for up to 2 years.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
Uracil DNA glycosylase	1 U
<i>Top</i> DNA Polymerase	1 U
dNTPs with dUTP	Each 200 µM
Reaction buffer with 1.5 mM MgCl ₂	1X
Stabilizer and tracking dye	O

Specifications

<i>Top</i> DNA Polymerase	
5' to 3' exonuclease activity	No
3' to 5' exonuclease activity	No
Terminal transferase activity	Yes
Fragment size	Up to 10 kb

Enzyme Inactivation

UDG is inactivated by heating at 95°C for 5 min.

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Precautions

- UDG activities can be remained after finishing reactions, if it is kept on below 50°C. Therefore, reaction mixture is recommended to freeze immediately after the reaction.

Online Resources



Korean



English

Visit our [product page](#) for additional information and protocols

Ordering Information

Description	Cat. No.		
0.2 ml thin-wall 8-tube	96 tubes	20 µl/rxn	K-2012-1
strips with attached cap	480 tubes	20 µl/rxn	K-2016-1



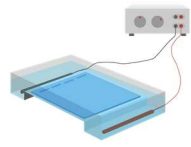

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols

Batch Code	Biological Risks	Catalog Number	Caution
Consult Instructions For Use	Contains Sufficient for <n> tests	Do not Re-use	Manufacturer
Research Use Only	Temperature Limitation	Use-by Date	

Experimental Procedures

Steps		Procedure Details																												
1	 Preparation of reaction mixture	<p>1. Add template DNA, primers, and nuclease-free water into <i>AccuPower®</i> PCR PreMix (with UDG) tubes to make a total volume of 20 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Amount of template <table border="1"> <thead> <tr> <th>Template DNA</th> <th>Amount of template</th> </tr> </thead> <tbody> <tr> <td>Bacteriophage λ, Plasmid DNA</td> <td>100 fg-200 ng</td> </tr> <tr> <td>Total genomic DNA</td> <td>1-500 ng</td> </tr> </tbody> </table> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1"> <thead> <tr> <th>Components</th> <th>20 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA</td> <td>Variable (1-10 µl)</td> </tr> <tr> <td>Forward primer (10 pmol/µl)</td> <td>0.5-2 µl</td> </tr> <tr> <td>Reverse primer (10 pmol/µl)</td> <td>0.5-2 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td>Variable</td> </tr> <tr> <td>Total volume</td> <td>20 µl</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried blue pellet by vortexing or pipetting, and briefly spin down.</p>	Template DNA	Amount of template	Bacteriophage λ, Plasmid DNA	100 fg-200 ng	Total genomic DNA	1-500 ng	Components	20 µl reaction	Template DNA	Variable (1-10 µl)	Forward primer (10 pmol/µl)	0.5-2 µl	Reverse primer (10 pmol/µl)	0.5-2 µl	Nuclease-free water	Variable	Total volume	20 µl										
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2	 Incubate reactions in a thermal cycler	<p>3. Perform the reaction under the following conditions.</p> <table border="1"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>UDG activation</td> <td>37°C</td> <td>2 min</td> <td>1 cycle</td> </tr> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>5 min</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>0.5-1 min</td> <td></td> </tr> <tr> <td>Annealing</td> <td>45-65°C*</td> <td>0.5-1 min</td> <td>25-35 cycles</td> </tr> <tr> <td>Extension</td> <td>72°C</td> <td>0.5-1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td>72°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> </tbody> </table> <p>* Set the annealing temperature to 3-5 degrees lower than the Tm of the primers</p>	Step	Temperature	Time	Cycles	UDG activation	37°C	2 min	1 cycle	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	0.5-1 min		Annealing	45-65°C*	0.5-1 min	25-35 cycles	Extension	72°C	0.5-1 min/kb		Final extension	72°C	3-5 min	1 cycle
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3	 Analyze with gel electrophoresis	<p>4. After the reaction, maintain the reaction mixture at -20°C or load samples on agarose gel immediately.</p> <p>5. Load samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																												
	 Option	<ul style="list-style-type: none"> If primer's Tm value is more than 65°C or PCR product size is more than 5 kb, follow the conditions as below. <table border="1"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>UDG activation</td> <td>37°C</td> <td>2 min</td> <td>1 cycle</td> </tr> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>5 min</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>30 sec</td> <td></td> </tr> <tr> <td>Annealing/Extension</td> <td>68°C</td> <td>1 min/kb</td> <td>30-35 cycles</td> </tr> <tr> <td>Final extension</td> <td>72°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> </tbody> </table>	Step	Temperature	Time	Cycles	UDG activation	37°C	2 min	1 cycle	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	30 sec		Annealing/Extension	68°C	1 min/kb	30-35 cycles	Final extension	72°C	3-5 min	1 cycle				
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