

[Cat. No.] **K-2909**

## Introduction

AccuPower® Clostridium perfringens PCR Kit is a ready-to-use premix for multiplex PCR that can be used to detect infection of *Clostridium perfringens* (*C. perfringens*).

*C. perfringens* is a gram-positive, anaerobic rod-shaped pathogenic bacterium. It is a major causative agent of bacterial food poisoning along with *Salmonella* and *Staphylococcus aureus*. When infected, *C. perfringens* bacteria produce A-E toxins and oval-shaped spores. Therefore, quantitative methods are mainly used to detect the five toxins. This product contains vacuum-dried components specific to *C. perfringens* including DNA polymerase, primers, dNTPs, and reaction buffer required for PCR. This ready-to-use kit simplifies preparation of PCR mixture as the user only has to add template DNA and nuclease-free water. After the reaction, since tracking dye is included, the samples can be applied directly on agarose gel for analysis without adding extra solution.

## Features & Benefits

- **Convenience & Reproducibility:** All reactants necessary for PCR including primers are lyophilized in each PCR tube, providing reproducible results in a convenient way.
- **Multiple amplification:** Generates three multiplex amplification products using only a single tube.
- **Sensitivity:** By applying the patented PyroHotStart (Enzyme-mediated HotStart) technology that minimizes non-specific reactions and maximizes reaction efficiency, only the target gene can be effectively amplified even with a trace amount of template DNA.
- **Stability:** Included stabilizer in the PCR reaction mixture provides increased stability compared to solution-type products.

## Composition

Composition	20 µl reaction
Top DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction buffer with 2 mM MgCl <sub>2</sub>	1X
Stabilizer and tracking dye	O
<i>C. perfringens</i> α Forward/Reverse primer	0.25 µM
<i>C. perfringens</i> β Forward/Reverse primer	0.25 µM
<i>C. perfringens</i> ε Forward/Reverse primer	0.25 µM
<i>C. perfringens</i> ι Forward/Reverse primer	0.25 µM

\* **Note:** For research use only. Not for use in diagnostic or therapeutic procedures.

## Specifications

Top DNA Polymerase	
5'→3' exonuclease activity	No
3'→5' exonuclease activity	No
3'-A overhang	Yes
Fragment size	α : 402 bp
	β : 236 bp
	ε : 541 bp
	ι : 317 bp

## Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

## Online Resources



English

Visit our **product page** for additional information and protocols

## Ordering Information

Description	Cat. No.
AccuPower® Clostridium perfringens PCR Kit, 0.2 ml thin-wall 8-tube strips with attached cap / 96 tubes	K-2909

## Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

## Explanation of Symbols



Batch Code



Biological Risks



Catalog Number



Caution



Consult Instructions For Use



Contains Sufficient for <n> tests



Do not Re-use



Manufacturer



Research Use Only




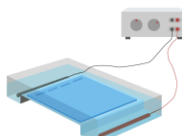


Temperature Limitation



Use-by Date

## Experimental Procedures

Steps		Procedure Details																								
1	<div></div> <div>Add template DNA</div>	<p>1. After preparing the template DNA and nuclease-free water, add the template DNA to the <i>AccuPower</i><sup>®</sup> <i>Clostridium perfringens</i> PCR Kit.</p>																								
2	<div></div> <div>Preparation of reaction mixture</div>	<p>2. Add nuclease-free water into PCR tubes to make a total volume of 20 µl. (Do not include the volume of the dried premix in the PCR tubes.)</p> <p>3. Completely dissolve the vacuum-dried pellet by vortexing, and briefly spin down.</p>																								
3	<div></div> <div>Incubate reactions in a thermal cycler</div>	<p>4. Place PCR tubes on the thermal cycler.</p> <p>5. Perform the reaction under the following conditions.</p> <table><thead><tr><th>Step</th><th>Temperature</th><th>Time</th><th>Cycles</th></tr></thead><tbody><tr><td>Pre-denaturation</td><td>92°C</td><td>5 min</td><td>1 cycle</td></tr><tr><td>Denaturation</td><td>92°C</td><td>30 sec</td><td></td></tr><tr><td>Annealing</td><td>57°C</td><td>30 sec</td><td>45 cycles</td></tr><tr><td>Extension</td><td>72°C</td><td>30 sec</td><td></td></tr><tr><td>Final extension</td><td>72°C</td><td>10 min</td><td>1 cycle</td></tr></tbody></table> <p>* <b>Note:</b> Users can adjust the protocol according to their instrument and template sequences to get optimal results.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	92°C	5 min	1 cycle	Denaturation	92°C	30 sec		Annealing	57°C	30 sec	45 cycles	Extension	72°C	30 sec		Final extension	72°C	10 min	1 cycle
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4	<div></div> <div>Analyze with gel electrophoresis</div>	<p>6. After the reaction, maintain the reaction mixture at 4-8°C.</p> <p>7. Load samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																								