

[Cat. No.] Please refer to the **Ordering Information**

Introduction

AccuPower® HotStart Pfu PCR PreMix is based on a concept chemical interaction between pyrophosphate (PPi) and pyrophosphatase (PPase). DNA polymerase is required Mg²⁺ to activate, but included PPi binds with high affinities to Mg²⁺ resulting in inhibition of polymerase activity. When the temperature rises during denaturation step, Mg-PPi complex is decomposed into 2Pi and Mg²⁺ by PPase. Then, activated DNA polymerase proceed reactions. This prevents the formation of mis-primed products and primer-dimers during the reaction set up process resulting in improved PCR specificity. Included Pfu DNA polymerase provide highly accurate PCR products through its proof-reading function. This product contains vacuum-dried components including Pfu DNA Polymerase, dNTPs, reaction buffer, stabilizer, and tracking dye. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, samples can be applied directly on agarose gel for analysis.

Applications

- Gene cloning with blunt ends
- Site-directed mutagenesis
- High fidelity amplification
- High specificity PCR
- cDNA amplification

Features & Benefits

- High fidelity: Low mutation rate during DNA amplification due to its high fidelity (error rate = 1.9×10^{-6}).
- Specificity: Minimized non-specific amplification and maximized PCR efficiency by using BIONEER's differentiated PyroHotStart technology.
- User-friendly: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- Stability: Included stabilizer enables delivery at room temperature and provides increased stability compared to solution-type products.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
Pfu DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction Buffer with 1.5 mM MgCl ₂	1X
Stabilizer and tracking dye	0

Specifications

Pfu DNA Polymerase	
5' to 3' exonuclease activity	No
3' to 5' exonuclease activity	Yes
3'-A overhang	No
Fragment size	Up to 5 kb

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

Ordering Information

Description	Cat. No.
0.2 ml thin-wall 8-tube strips with attached cap	96 tubes 20 µl/rxn K-2301
	50 µl/rxn K-2302
480 tubes	20 µl/rxn K-2303
	50 µl/rxn K-2304

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Batch Code



Biological Risks



Catalog Number



Caution



Consult Instructions For Use



Contains Sufficient for <n> tests



Do not Re-use



Manufacturer



Research Use Only



Temperature Limitation



Use-by Date

Experimental Procedures

Steps		Procedure Details																								
1	 Preparation of reaction mixture	<p>1. Add template DNA, primers, and nuclease-free water into <i>AccuPower® HotStart Pfu PCR PreMix</i> tubes to make a total volume of 20 µl or 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Components</th> <th style="text-align: center;">20 µl reaction</th> <th style="text-align: center;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA (1-500 ng)</td> <td style="text-align: center;">Variable (1-10 µl)</td> <td style="text-align: center;">Variable (1-25 µl)</td> </tr> <tr> <td>Forward primer (10 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Reverse primer (10 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td style="text-align: center;">Variable</td> <td style="text-align: center;">Variable</td> </tr> <tr> <td>Total volume</td> <td style="text-align: center;">20 µl</td> <td style="text-align: center;">50 µl</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried blue pellet by vortexing or pipetting, and briefly spin down.</p>	Components	20 µl reaction	50 µl reaction	Template DNA (1-500 ng)	Variable (1-10 µl)	Variable (1-25 µl)	Forward primer (10 pmol/µl)	0.5-2 µl	1-5 µl	Reverse primer (10 pmol/µl)	0.5-2 µl	1-5 µl	Nuclease-free water	Variable	Variable	Total volume	20 µl	50 µl						
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2	 Incubate reactions in a thermal cycler	<p>3. Perform the reaction under the following conditions.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">30 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">55°C</td> <td style="text-align: center;">30 sec</td> <td style="text-align: center;">30-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* Note: For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	30 sec		Annealing	55°C	30 sec	30-35 cycles	Extension	72°C	1 min/kb		Final extension	72°C	5 min	1 cycle
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3	 Analyze with gel electrophoresis	<p>4. After the reaction, maintain the reaction mixture at 4-8°C. The samples can be stored at -20°C until use.</p> <p>5. Load 5 µl of samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																								