

[Cat. No.] **K-6400**

Introduction

AccuPower® GreenStar™ RT-qPCR PreMix is a one-step RT-qPCR product based on intercalating dye method using thermostable reverse transcriptase (RTase) and HotStart PCR technology. By applying BIONEER's RocketScript™ RTase which is enhanced thermal stability and processivity, efficient reverse transcription (RT) of RNA molecules with complex secondary structures is possible. In real-time PCR, antibody-based HotStart Taq DNA Polymerase provides superior priming accuracy and reduced non-specific reactions such as mis-priming and primer dimer during PCR at a low temperature. This product contains vacuum-dried all components for real-time PCR, except for template RNA and target-specific primers. By just adding template RNA and target-specific primers, reproducible results with high sensitivity and specificity can be obtained.

Applications

- Low copy viral/bacterial pathogen load determination in an earlier stage
- Low copy mRNA amplification
- Low copy target RNA quantification
- RNA amplification for microarray and/or for NGS

Features & Benefits

- High sensitivity: Amplification of target gene present in a miniscule amount of 1 pg template RNA.
- High specificity: Minimized experimental errors by non-specific amplification and effective amplification of template RNA existing in a small amount by using HotStart Taq DNA Polymerase and thermostable RTase.
- Advanced performance: Comprehensive choice of template RNA for RT-qPCR, even complex secondary structure, by using thermostable RocketScript™ RTase, capable of performing RT at high temperature.
- Convenience: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform one-step RT-qPCR by adding template RNA and target-specific primers.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Components

| Components | K-6400 |
|------------|---------------|
| Tube/Plate | 96 tubes |
| DEPC-D.W. | 1.8 ml x 4 ea |

Composition

| Composition | Concentration |
|---|---------------|
| HotStart Taq DNA Polymerase | 1 U |
| RocketScript Reverse Transcriptase | 200 U |
| Intercalating dye | 0.4X |
| Reaction buffer with 1.5 mM MgCl ₂ | 1X |
| dNTPs (dATP, dCTP, dGTP, dTTP) | 1 mM |

Specifications

| HotStart Taq DNA Polymerase | |
|-------------------------------|-----|
| 5' to 3' exonuclease activity | Yes |
| 3' to 5' exonuclease activity | No |
| 3'-A overhang | Yes |

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

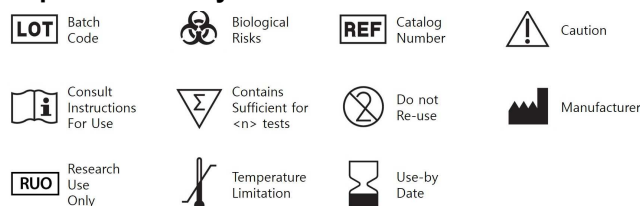
Ordering Information

| Description | | | | | Cat. No. |
|-------------|---------------|-------|-----------------------|--------|----------|
| Exicycler | 8-tube strips | 50 µl | optical film included | 96 rxn | K-6400 |



Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Experimental Procedures

| Steps | Procedure Details | | | | | | | | | | | | | | | | | | | |
|---|--|------------|----------------|-----------------------------|----------|-----------------------------|----------|-----------------------------|----------|-------------------------|------|-----------|----------|--------------|-------|----------|--------------|-----------------------|---------|----------|
| <p style="text-align: center;">1</p> <div style="text-align: center;">  <p>Preparation of reaction mixture</p> </div> | <p>1. Add template RNA, target-specific primers, 50X ROX dye (optional, not provided), and DEPC-D.W. into <i>AccuPower® GreenStar™</i> RT-qPCR PreMix tubes to make a total volume of 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 70%;">Components</th> <th style="width: 30%;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template RNA (10 pg-100 ng)</td> <td>Variable</td> </tr> <tr> <td>Forward primer (10 pmol/µl)</td> <td>0.5-3 µl</td> </tr> <tr> <td>Reverse primer (10 pmol/µl)</td> <td>0.5-3 µl</td> </tr> <tr> <td>(Optional) 50X ROX dye*</td> <td>1 µl</td> </tr> <tr> <td>DEPC-D.W.</td> <td>Variable</td> </tr> <tr> <td>Total volume</td> <td>50 µl</td> </tr> </tbody> </table> <p>* Note: ROX dye is used for normalization of intensity by background subtraction. The use of ROX dye is recommended for Applied Biosystems 7500 Real-Time PCR System (Applied Biosystems), but not required for <i>Exicycler™</i> 96 Real-Time PCR System (BIONEER) and CFX96 Real-Time PCR System (Bio-Rad).</p> <p>2. Seal real-time PCR tubes with adhesive optical sealing film (Cat. No. 3111-4110, provided).</p> <p>3. Dissolve the vacuum-dried pellet by vortexing, and briefly spin down.</p> | Components | 50 µl reaction | Template RNA (10 pg-100 ng) | Variable | Forward primer (10 pmol/µl) | 0.5-3 µl | Reverse primer (10 pmol/µl) | 0.5-3 µl | (Optional) 50X ROX dye* | 1 µl | DEPC-D.W. | Variable | Total volume | 50 µl | | | | | |
| Components | 50 µl reaction | | | | | | | | | | | | | | | | | | | |
| Template RNA (10 pg-100 ng) | Variable | | | | | | | | | | | | | | | | | | | |
| Forward primer (10 pmol/µl) | 0.5-3 µl | | | | | | | | | | | | | | | | | | | |
| Reverse primer (10 pmol/µl) | 0.5-3 µl | | | | | | | | | | | | | | | | | | | |
| (Optional) 50X ROX dye* | 1 µl | | | | | | | | | | | | | | | | | | | |
| DEPC-D.W. | Variable | | | | | | | | | | | | | | | | | | | |
| Total volume | 50 µl | | | | | | | | | | | | | | | | | | | |
| <p style="text-align: center;">2</p> <div style="text-align: center;">  <p>RT-qPCR</p> </div> | <p>4. Perform the reaction under the following conditions.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 25%;">Step</th> <th style="width: 25%;">Temperature</th> <th style="width: 25%;">Time</th> <th style="width: 25%;">Cycles</th> </tr> </thead> <tbody> <tr> <td>cDNA synthesis</td> <td>50-70°C†</td> <td>15 min</td> <td>1 cycle</td> </tr> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>5-30 sec</td> <td rowspan="2">40-45 cycles</td> </tr> <tr> <td>Annealing & Extension</td> <td>55-60°C</td> <td>5-30 sec</td> </tr> </tbody> </table> <p>* Note: Users can adjust the protocol according to their instrument and template sequences to get optimal results.</p> <p>† For cDNA synthesis, it is recommended to start your reaction at least at 50°C, but not exceed at 70°C.</p> <p>5. After the reaction is completed, analyze the results.</p> | Step | Temperature | Time | Cycles | cDNA synthesis | 50-70°C† | 15 min | 1 cycle | Pre-denaturation | 95°C | 3-5 min | 1 cycle | Denaturation | 95°C | 5-30 sec | 40-45 cycles | Annealing & Extension | 55-60°C | 5-30 sec |
| Step | Temperature | Time | Cycles | | | | | | | | | | | | | | | | | |
| cDNA synthesis | 50-70°C† | 15 min | 1 cycle | | | | | | | | | | | | | | | | | |
| Pre-denaturation | 95°C | 3-5 min | 1 cycle | | | | | | | | | | | | | | | | | |
| Denaturation | 95°C | 5-30 sec | 40-45 cycles | | | | | | | | | | | | | | | | | |
| Annealing & Extension | 55-60°C | 5-30 sec | | | | | | | | | | | | | | | | | | |