

[Cat. No.] Please refer to the **Ordering Information**

Introduction

AccuPower® Gold Multiplex PCR PreMix is the powerful technology for convenient and easy performance that allows DNA amplification of two or more products in a single tube. It is based on a concept chemical interaction between pyrophosphate (PPi) and pyrophosphatase (PPase). Included PPi binds with high affinities to Mg²⁺ resulting in inhibition of polymerase activity. When the temperature rises during denaturation step, Mg-PPi complex is decomposed into 2Pi and Mg²⁺ by PPase. Then, activated DNA polymerase proceed reactions. This prevents the formation of mis-primed products and primer-dimers during the reaction set up process resulting in improved PCR specificity. This product contains vacuum-dried components including HotStart DNA Polymerase, dNTPs, reaction buffer, stabilizer, and tracking dye. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, samples can be applied directly on agarose gel for analysis.

Applications

- STR analysis
- Molecular diagnostic analysis
- Qualitative, semi-qualitative gene expression assay
- Mutant screening
- Transgenic organism analysis
- Genotyping assay

Features & Benefits

- Multiplex PCR: Generation of 20 multiplexed amplification products in a single tube.
- Specificity & Efficiency: Minimized non-specific amplification and maximized PCR efficiency by using HotStart DNA Polymerase.
- Sensitivity: Excellent sensitivity and amplification efficiency even with small amounts of DNA.
- User-friendly: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- Stability: Included stabilizer enables delivery at room temperature and provides increased stability compared to solution-type products.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
HotStart DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction buffer with 2 mM MgCl ₂	1X
Stabilizer and tracking dye	○

Specifications

HotStart DNA Polymerase	
5' to 3' exonuclease activity	No
3' to 5' exonuclease activity	No
3'-A overhang	Yes
Fragment size	Up to 1 kb

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Precautions

- This enzyme is specifically optimized for increasing base incorporation rate by inactivating 5' to 3' exonuclease activity. Therefore, this product is not recommended to use for real-time PCR using hydrolysis probe method.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

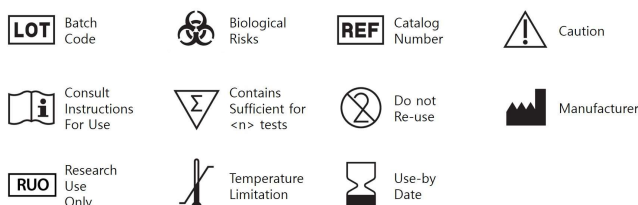
Ordering Information

Description	Cat. No.
0.2 ml thin-wall 8-tube strips with attached cap	96 tubes 20 µl/rxn K-2115
	50 µl/rxn K-2117
480 tubes	20 µl/rxn K-2116
	50 µl/rxn K-2118



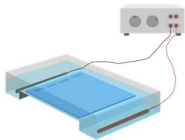
Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Experimental Procedures

Steps		Procedure Details																								
1	 Preparation of reaction mixture	<p>1. Add template DNA, primers, and nuclease-free water into <i>AccuPower®</i> Gold Multiplex PCR PreMix tubes to make a total volume of 20 µl or 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Components</th> <th style="text-align: center;">20 µl reaction</th> <th style="text-align: center;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA (1-100 ng)</td> <td style="text-align: center;">Variable (1-10 µl)</td> <td style="text-align: center;">Variable (1-25 µl)</td> </tr> <tr> <td>Forward primer (1-5 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Reverse primer (1-5 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td style="text-align: center;">Variable</td> <td style="text-align: center;">Variable</td> </tr> <tr> <td>Total volume</td> <td style="text-align: center;">20 µl</td> <td style="text-align: center;">50 µl</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried green pellet by vortexing or pipetting, and briefly spin down.</p>	Components	20 µl reaction	50 µl reaction	Template DNA (1-100 ng)	Variable (1-10 µl)	Variable (1-25 µl)	Forward primer (1-5 pmol/µl)	0.5-2 µl	1-5 µl	Reverse primer (1-5 pmol/µl)	0.5-2 µl	1-5 µl	Nuclease-free water	Variable	Variable	Total volume	20 µl	50 µl						
Components	20 µl reaction	50 µl reaction																								
Template DNA (1-100 ng)	Variable (1-10 µl)	Variable (1-25 µl)																								
Forward primer (1-5 pmol/µl)	0.5-2 µl	1-5 µl																								
Reverse primer (1-5 pmol/µl)	0.5-2 µl	1-5 µl																								
Nuclease-free water	Variable	Variable																								
Total volume	20 µl	50 µl																								
2	 Incubate reactions in a thermal cycler	<p>3. Perform the reaction under the following conditions.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">30 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">55-65°C[†]</td> <td style="text-align: center;">30-60 sec</td> <td style="text-align: center;">25-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* Note: Primers are generally designed length of 24-35 nucleotides and ideally have a T_m value range within 5°C. [†] Set the annealing temperature to 3-5 degrees lower than the T_m of the primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	30 sec		Annealing	55-65°C [†]	30-60 sec	25-35 cycles	Extension	72°C	1 min/kb		Final extension	72°C	5 min	1 cycle
Step	Temperature	Time	Cycles																							
Pre-denaturation	95°C	5 min	1 cycle																							
Denaturation	95°C	30 sec																								
Annealing	55-65°C [†]	30-60 sec	25-35 cycles																							
Extension	72°C	1 min/kb																								
Final extension	72°C	5 min	1 cycle																							
3	 Analyze with gel electrophoresis	<p>4. After the reaction, maintain the reaction mixture at 4-8°C. The samples can be stored at -20°C until use.</p> <p>5. Load 5 µl of samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																								