

[Cat. No.] Please refer to the **Ordering Information**

### Introduction

AccuPower® CycleScript™ RT PreMix (dN<sub>12</sub>) is applied with BIONEER's patent technology called Cyclic Temperature Reverse Transcription (CTRT), which not only increases the efficiency, but also is effective for full-length cDNA synthesis. The CTRT reaction is composed of 2 or 3 steps. The step 1 is performed at 15-25°C, at which short primers are fully annealed. Then, the step 2 is performed at 42-48°C for cDNA synthesis. The step 3 is performed at high temperature 50-55°C at which secondary structure of template RNA obstructing reverse transcription is released. This product contains vacuum-dried components including thermostable CycleScript™ Reverse Transcriptase, reaction buffer, DTT, dNTPs, Oligo dN<sub>12</sub> primers, and stabilizer. It simplifies preparation of reverse transcription reaction mixture by adding template RNA and nuclease-free water without any extra process.

### Applications

- Sequencing single and double-strand DNA or RNA
- Random priming reaction
- cDNA library construction
- Probe labeling
- mRNA 5'-end mapping by primer extension analysis
- PCR
- Real-time PCR

### Features & Benefits

- Flexible conditions: Cyclic Temperature Reverse Transcription (CTRT) is more efficient cDNA synthesis by using higher temperature than the conventional methods using at 42°C. Primer annealing is performed at a range of low temperature (15-40°C) and secondary structures of template is released by repeating the step 2 or 3 at a range of high temperature (50-55°C).
- Stability: Included stabilizer and thermostable reverse transcriptase allow to react at a higher temperature up to 55°C.
- Controllable reaction time: Reaction time can be controlled depending on the number and size of copies of target gene. In case of high-copy gene, cDNA can be synthesized even with 10 minutes reverse transcription reaction.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.
- Ease-of-use: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform cDNA synthesis by adding template RNA and nuclease-free water.

### Composition

Composition	Concentration
CycleScript™ Reverse Transcriptase	200 U
5X Reaction buffer	1X
DTT	0.25 mM
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
RNase inhibitors	1 U
Oligo dN <sub>12</sub>	100 pmol

### Specifications

CycleScript™ Reverse Transcriptase	
DNase activity	No
RNase activity	No
Fragment size	Up to 9 kb

### Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

### Online Resources



Korean



English

Visit our **product page** for additional information and protocols

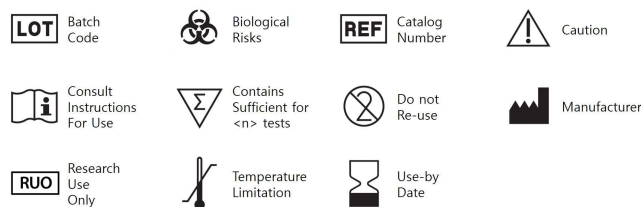
### Ordering Information

Description	Cat. No.
0.2 ml thin-wall 96 tubes	K-2045
8-tube strips with attached cap	K-2048
480 tubes	K-2045-B
50 µl/rxn	K-2048-B
0.5 ml thin-wall tubes with attached cap	K-2050-1




### Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

### Explanation of Symbols



**Experimental Procedures**

Steps		Procedure Details																																																	
1	 <b>Preparation of reaction mixture</b>	<p>1. Add template RNA and nuclease-free water into <i>AccuPower® CycleScript™ RT PreMix (dN<sub>12</sub>)</i> tubes to make a total volume of 20 µl or 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> <li>Amount of template RNA</li> </ul> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th colspan="2">Components</th> <th>20 µl reaction</th> <th colspan="2">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td rowspan="2" style="text-align: center;">Template RNA</td> <td style="text-align: center;">Total RNA</td> <td style="text-align: center;">0.1-1.0 µg</td> <td colspan="2" style="text-align: center;">0.1-1.0 µg</td> </tr> <tr> <td style="text-align: center;">Poly(A) RNA</td> <td style="text-align: center;">0.01-1.0 µg</td> <td colspan="2" style="text-align: center;">0.01-1.0 µg</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried pellet by pipetting or vortexing, and briefly spin down.</p>				Components		20 µl reaction	50 µl reaction		Template RNA	Total RNA	0.1-1.0 µg	0.1-1.0 µg		Poly(A) RNA	0.01-1.0 µg	0.01-1.0 µg																																	
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2	 <b>cDNA synthesis</b>	<p>3. Perform the reaction under the following conditions.</p> <p>3-1. CTRT reaction (Example 1)</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Primer annealing</td> <td style="text-align: center;">15-25°C</td> <td style="text-align: center;">30 sec</td> <td rowspan="2" style="text-align: center;">12 cycles or less</td> </tr> <tr> <td>cDNA synthesis</td> <td style="text-align: center;">42-45°C</td> <td style="text-align: center;">4 min</td> </tr> <tr> <td>Melting secondary structure &amp; cDNA synthesis</td> <td style="text-align: center;">55°C</td> <td style="text-align: center;">30 sec</td> <td></td> </tr> <tr> <td>Heat inactivation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>3-2. CTRT reaction (Example 2)</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Primer annealing</td> <td style="text-align: center;">15-25°C</td> <td style="text-align: center;">1 min</td> <td rowspan="2" style="text-align: center;">12 cycles or less</td> </tr> <tr> <td>Melting secondary structure &amp; cDNA synthesis</td> <td style="text-align: center;">42-50°C</td> <td style="text-align: center;">4 min</td> </tr> <tr> <td>Heat inactivation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>3-3. Single temperature reaction (Example 3)</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>cDNA synthesis</td> <td style="text-align: center;">37-50°C</td> <td style="text-align: center;">30-60 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Heat inactivation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* <b>Note:</b> Recommended temperature is range of 42-48°C.</p> <p>4. After the reaction, maintain the reaction mixture at 4°C. The samples can be stored at -20°C until use.</p>				Step	Temperature	Time	Cycles	Primer annealing	15-25°C	30 sec	12 cycles or less	cDNA synthesis	42-45°C	4 min	Melting secondary structure & cDNA synthesis	55°C	30 sec		Heat inactivation	95°C	5 min	1 cycle	Step	Temperature	Time	Cycles	Primer annealing	15-25°C	1 min	12 cycles or less	Melting secondary structure & cDNA synthesis	42-50°C	4 min	Heat inactivation	95°C	5 min	1 cycle	Step	Temperature	Time	Cycles	cDNA synthesis	37-50°C	30-60 min	1 cycle	Heat inactivation	95°C	5 min	1 cycle
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	 <b>Option</b>	<ul style="list-style-type: none"> <li>If PCR is needed, transfer 2-5 µl of reaction mixture (synthesized cDNA) into <i>AccuPower® PCR PreMix</i> tubes (Cat. No. K-2012, not provided), and perform the reaction under the following conditions.</li> </ul> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">20 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">45-65°C</td> <td style="text-align: center;">20 sec</td> <td style="text-align: center;">25-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">0.5-1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">3-5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* <b>Note:</b> For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers.</p>				Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	20 sec		Annealing	45-65°C	20 sec	25-35 cycles	Extension	72°C	0.5-1 min/kb		Final extension	72°C	3-5 min	1 cycle																						
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