

[Cat. No.] **K-2051**

## Introduction

AccuPower® CycleScript™ RT Master Mix is applied with BIONEER's patent technology called Cyclic Temperature Reverse Transcription (CTRT), which not only increases the efficiency, but also is effective for full-length cDNA synthesis. The CTRT reaction is composed of 2 or 3 steps. The step 1 is performed at 15-25°C, at which short primers are fully annealed. Then, the step 2 is performed at 42-48°C for cDNA synthesis. The step 3 is performed at high temperature 50-55°C at which secondary structure of template RNA obstructing reverse transcription is released. This product is a ready-to-use mixture containing thermostable CycleScript™ Reverse Transcriptase, reaction buffer, DTT, dNTPs, and stabilizer. Primers (Oligo dT<sub>20</sub> and Oligo dN<sub>6</sub>) are provided in separated tubes. It simplifies preparation of reverse transcription reaction mixture by adding template RNA, primers, and nuclease-free water without any extra process.

## Applications

- Sequencing single and double-strand DNA or RNA
- Random priming reaction
- cDNA library construction
- Probe labeling
- mRNA 5'-end mapping by primer extension analysis
- PCR
- Real-time PCR

## Features & Benefits

- Flexible conditions: Cyclic Temperature Reverse Transcription (CTRT) is more efficient cDNA synthesis by using higher temperature than the conventional methods using at 42°C. Primer annealing is performed at a range of low temperature (15-40°C) and secondary structures of template is released by repeating the step 2 or 3 at a range of high temperature (50-55°C).
- Stability: Included stabilizer and thermostable reverse transcriptase allow to react at a higher temperature up to 55°C.
- Controllable reaction time: Reaction time can be controlled depending on the number and size of copies of target gene. In case of high-copy gene, cDNA can be synthesized even with 10 minutes reverse transcription reaction.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.
- Ease-of-use: Reactants are included in a tube, it allows any user simply perform cDNA synthesis by adding template RNA and nuclease-free water.

## Components

Components	Amount
2X Master Mix	1 ml
Oligo dT <sub>20</sub> (100 pmol/μl)	100 μl
Oligo dN <sub>6</sub> (100 pmol/μl)	100 μl

## Composition

Composition	Concentration
CycleScript™ Reverse Transcriptase	200 U
5X Reaction buffer	1X
DTT	0.25 mM
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 μM
RNase inhibitors	1 U

## Specifications

CycleScript™ Reverse Transcriptase	
DNase activity	No
RNase activity	No
Fragment size	Up to 9 kb

## Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

## Online Resources



Korean



English

Visit our **product page** for additional information and protocols

## Ordering Information

Description	Cat. No.
1 ml of 2X Master Mix solution	1 ml x 1 ea K-2051

\* **Note:** For Master Mix products, primers (dT<sub>20</sub> & dN<sub>6</sub>) are provided in a separate tube.

## Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

## Explanation of Symbols



Batch Code



Biological Risks



Catalog Number



Caution



Consult Instructions For Use



Contains Sufficient for <n> tests



Do not Re-use



Manufacturer



Research Use Only







Temperature Limitation



Use-by Date

## Experimental Procedures

Steps		Procedure Details																																															
1	<div></div> <div>Thaw reagents</div>	<div>1. Thaw <i>AccuPower® CycleScript™</i> RT Master Mix on ice and mix thoroughly before use. Then, briefly spin down components.</div> <div>2. Dispense appropriate volumes of <i>AccuPower® CycleScript™</i> RT Master Mix into PCR tubes (not provided). Use 10 µl and 25 µl of 2X Master Mix for 20 µl reaction and 50 µl reaction, respectively.</div>																																															
2	<div></div> <div>Preparation of reaction mixture</div>	<div>3. Add template RNA, primers, and nuclease-free water into PCR tubes containing <i>AccuPower® CycleScript™</i> RT Master Mix.</div> <div>• Amount of template RNA and primers</div> <table><tr><th colspan="2">Components</th><th>20 µl reaction</th><th>50 µl reaction</th></tr><tr><td rowspan="2">Template RNA</td><td>Total RNA</td><td>0.1-1.0 µg</td><td>0.1-1.0 µg</td></tr><tr><td>Poly(A) RNA</td><td>0.01-1.0 µg</td><td>0.01-1.0 µg</td></tr><tr><td rowspan="3">Primers</td><td>Oligo dT</td><td>50-100 pmol</td><td>250 pmol</td></tr><tr><td>Random primer</td><td>100 pmol</td><td>250 pmol</td></tr><tr><td>Gene specific primer</td><td>10-50 pmol</td><td>10-50 pmol</td></tr></table> <div>4. Mix the reaction mixture by pipetting or vortexing, and briefly spin down.</div>			Components		20 µl reaction	50 µl reaction	Template RNA	Total RNA	0.1-1.0 µg	0.1-1.0 µg	Poly(A) RNA	0.01-1.0 µg	0.01-1.0 µg	Primers	Oligo dT	50-100 pmol	250 pmol	Random primer	100 pmol	250 pmol	Gene specific primer	10-50 pmol	10-50 pmol																								
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3	<div></div> <div>cDNA synthesis</div>	<div>5. Perform the reaction under the following conditions.</div> <div>5-1. CTRT reaction (Example 1)</div> <table><tr><th>Step</th><th>Temperature</th><th>Time</th><th>Cycles</th></tr><tr><td>Primer annealing</td><td>15-25°C</td><td>30 sec</td><td rowspan="3">12 cycles or less</td></tr><tr><td>cDNA synthesis</td><td>42-45°C</td><td>4 min</td></tr><tr><td>Melting secondary structure &amp; cDNA synthesis</td><td>55°C</td><td>30 sec</td></tr><tr><td>Heat inactivation</td><td>95°C</td><td>5 min</td><td>1 cycle</td></tr></table> <div>5-2. CTRT reaction (Example 2)</div> <table><tr><th>Step</th><th>Temperature</th><th>Time</th><th>Cycles</th></tr><tr><td>Primer annealing</td><td>15-25°C</td><td>1 min</td><td rowspan="2">12 cycles or less</td></tr><tr><td>Melting secondary structure &amp; cDNA synthesis</td><td>42-50°C</td><td>4 min</td></tr><tr><td>Heat inactivation</td><td>95°C</td><td>5 min</td><td>1 cycle</td></tr></table> <div>* <b>Note:</b> For difficult or high GC-content templates, perform cDNA synthesis at 55°C.</div> <div>5-3. Single temperature reaction (Example 3)</div> <table><tr><th>Step</th><th>Temperature</th><th>Time</th><th>Cycles</th></tr><tr><td>cDNA synthesis</td><td>37-50°C</td><td>30-60 min</td><td>1 cycle</td></tr><tr><td>Heat inactivation</td><td>95°C</td><td>5 min</td><td>1 cycle</td></tr></table> <div>* <b>Note:</b> Recommended temperature is range of 42-48°C.</div> <div>6. After the reaction, maintain the reaction mixture at 4°C. The samples can be stored at -20°C until use.</div>			Step	Temperature	Time	Cycles	Primer annealing	15-25°C	30 sec	12 cycles or less	cDNA synthesis	42-45°C	4 min	Melting secondary structure & cDNA synthesis	55°C	30 sec	Heat inactivation	95°C	5 min	1 cycle	Step	Temperature	Time	Cycles	Primer annealing	15-25°C	1 min	12 cycles or less	Melting secondary structure & cDNA synthesis	42-50°C	4 min	Heat inactivation	95°C	5 min	1 cycle	Step	Temperature	Time	Cycles	cDNA synthesis	37-50°C	30-60 min	1 cycle	Heat inactivation	95°C	5 min	1 cycle
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	<div></div> <div>Option</div>	<div>• If PCR is needed, transfer 2-5 µl of reaction mixture (synthesized cDNA) into <i>AccuPower®</i> PCR PreMix tubes (Cat. No. K-2012, not provided), and perform the reaction under the following conditions.</div> <table><tr><th>Step</th><th>Temperature</th><th>Time</th><th>Cycles</th></tr><tr><td>Pre-denaturation</td><td>95°C</td><td>5 min</td><td>1 cycle</td></tr><tr><td>Denaturation</td><td>95°C</td><td>20 sec</td><td rowspan="3">25-35 cycles</td></tr><tr><td>Annealing</td><td>45-65°C</td><td>20 sec</td></tr><tr><td>Extension</td><td>72°C</td><td>0.5-1 min/kb</td></tr><tr><td>Final extension</td><td>72°C</td><td>3-5 min</td><td>1 cycle</td></tr></table> <div>* <b>Note:</b> For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers.</div>			Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	20 sec	25-35 cycles	Annealing	45-65°C	20 sec	Extension	72°C	0.5-1 min/kb	Final extension	72°C	3-5 min	1 cycle																							
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