

## AccuPrep® Genomic DNA Extraction Kit (K-3032G)

### I Before You Begin

- 1) Completely dissolve one vial of **Proteinase K** in **1,250 µl** of nuclease-free water. For short term storage, dissolved Proteinase K should be stored at **4°C**. For long term storage, it is recommended to aliquot the enzyme into separate tubes and store at **-20°C**.
- 2) Completely dissolve one vial of **RNase A** in **600 µl** of nuclease-free water. For short term storage, dissolved RNase A should be stored at **4°C**. For long term storage, it is recommended to aliquot the enzyme into separate tubes and store at **-20°C**.
- 3) Add correct amount of **absolute ethanol (not provided)** to **WA1 Buffer**.
- 4) Add correct amount of **absolute ethanol (not provided)** to **WB2 Buffer**.
- 5) Pre-heat EA Buffer at **60°C** before use.

### II DNA Extraction from Whole Blood and Buffy Coat

- 1) Add **20 µl** of **Proteinase K** to a 1.5 ml or 2 ml tube.
- 2) Apply **200 µl** of **whole blood** or **buffy coat** to the tube containing Proteinase K.
- 3) Add **200 µl** of **GB Buffer** to the sample and mix well by vortexing.
- 4) Incubate at **60°C** for **10 min**.
- 5) Add **400 µl** of **absolute ethanol** (not provided) and mix well by pipetting.
- 6) Carefully transfer the lysate into the upper reservoir of the Binding column tube (fitted in a collection tube) without wetting the rim.
- 7) Close the tube and centrifuge at **8,000 rpm** for **1 min**.
- 8) Discard the solution from the collection tube and reuse the collection tube.
- 9) Add **500 µl** of **WA1 Buffer** without wetting the rim, close the tube, and centrifuge at **8,000 rpm** for **1 min**.
- 10) Discard the solution from the collection tube and reuse the collection tube.
- 11) Add **500 µl** of **WB2 Buffer** without wetting the rim, close the tube, and centrifuge at **8,000 rpm** for **1 min**.
- 12) Discard the solution from the collection tube and reuse the collection tube.
- 13) Centrifuge once more at **13,000 rpm** for **1 min** to completely remove ethanol, and check that there is no droplet clinging to the bottom of Binding column tube.
- 14) Transfer the Binding column tube to a new 1.5 ml tube for elution, add **50-200 µl** of **EA Buffer** onto Binding column tube, and wait for at least 1 min at RT (15-25°C).
- 15) Centrifuge at **8,000 rpm** for **1 min** to elute.

### III DNA Extraction from Cultured Cell

- 1) Centrifuge the cultured cells ( $10^4 \sim 10^6$  cells) for **5 min** at **300 x g**. Discard the supernatant carefully without disturbing the pellet.
- 2) Resuspend the pellet in **200 µl** of **1X PBS (not provided)**.
- 3) Add **20 µl** of **Proteinase K**.

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- 4) Add **10 µl** of **RNase A**, mix thoroughly and incubate the tubes for 2 min at room temperature.
- 5) Go to **step 3** of “DNA Extraction from Whole Blood and Buffy Coat” in page 1 and follow the instructions accordingly.

### IV DNA Extraction from Animal Tissue

- 1) Homogenize the sample (25-50 mg) with a mortar and pestle, place them in a clean 1.5 ml tube, and add **200 µl** of **TL Buffer**.
- 2) Add **20 µl** of **Proteinase K** and **10 µl** of **RNase A**. Mix by vortexing.
- 3) Incubate at **60°C** for **1 hour**, or until the tissue is completely lysed.
- 4) Add **200 µl** of **GB Buffer**, and mix by vortexing.
- 5) Go to **step 5** of “DNA Extraction from Whole Blood and Buffy Coat” in page 1 and continue the extraction process.

### V DNA Extraction from Gram-Negative Bacteria

- 1) Collect the bacterial cells up to  $\sim 1 \times 10^9$  cells by centrifuging at **8,000 rpm** for **5 min**. Discard the supernatant (media) by using a pipette.
- 2) Add **180 µl** of **TL Buffer** to the collected cell pellet and completely resuspend by vortexing or pipetting. Transfer the cell suspension to 1.5 ml or 2 ml tube.
- 3) Add **20 µl** of **Proteinase K** and **10 µl** of **RNase A**. Mix by vortexing.
- 4) Incubate at **60°C** for **1 hour**.
- 5) Add **200 µl** of **GB Buffer**, and immediately mix by vortex mixer.
- 6) Go to **step 5** of “DNA Extraction from Whole Blood and Buffy Coat” in page 1 and continue the extraction process.

### VI DNA Extraction from Gram-Positive Bacteria

- 1) Collect the bacterial cells by up to  $1 \times 10^9$  cells by centrifuging at **8,000 rpm** for **5 min**. Discard the supernatant (media) by using a pipette.
- 2) Add **180 µl** of **Lysis Buffer** for Gram-Positive bacteria (not provided) to the collected cell pellet and completely resuspend by vortexing or pipetting. Transfer the cell suspension to 1.5 ml or 2 ml tube.  
(Note) Lysis Buffer for Gram-Positive bacteria : 20 mM Tris-HCl (pH 8.0), 2 mM EDTA and 1.2% Triton® X-100.
- 3) Add **20 µl** of **lysozyme** (100 mg/ml, not provided) and **10 µl** of **RNase A** and mix thoroughly by vortexing.
- 4) Incubate the tubes at **37°C** for **30 min**.
- 5) Add **20 µl** of **Proteinase K**.
- 6) Add **200 µl** of **GB Buffer** and mix thoroughly by vortexing.
- 7) Incubate the tubes at **60°C** for **30 min** or until bacterial cells are completely lysed.
- 8) Go to **step 5** of “DNA Extraction from Whole Blood and Buffy Coat” in page 1 and continue the extraction process.

※ For more information, please visit [www.bioneer.com](http://www.bioneer.com) and refer to the User Guide of this kit.